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ULTRAVIOLET IRRADIATION TO ENRICH FOODS WITH VITAMIN D (review)

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Abstract

Vitamin D deficiency found in 50-90 % of the adult and children's population in Russia (I.N. Zakharova et al., 2015; V.M. Kodentsova et al., 2017, 2018) and caused by inadequate intake and reduced endogenous synthesis in the skin due to insufficient solar irradiation, is associated with many chronic diseases and makes an important problem (A. Hossein-nezhad et al., 2013). One of the options for biofortification, called "bio-addition", is based on the ability of living organisms to form vitamin D from endogenous ergosterol by UV irradiation. Ultraviolet irradiation of animals allows minimizing seasonal variations in the concentration of vitamin D in cow's milk (R.R. Weir et al., 2017). A one-hour exposure of animals for 14-day to insolation at summer noon increased the vitamin D₃ content in pork ($p < 0.001$) to $0.716 \pm 0.097 \mu\text{g}/100 \text{ g}$ ($28.6 \pm 3.9 \text{ IU}/100 \text{ g}$) which significantly exceeded the same indicator in the control animals ($0.218 \pm 0.024 \mu\text{g}/100 \text{ g}$, or $8.7 \pm 1.0 \text{ IU per } 100 \text{ g}$) (D.E. Larson-Meyer et al., 2017). UV irradiation effectively increased vitamin D level in chicken, from 0.16 to 0.96 $\mu\text{g per } 100 \text{ g}$, even at 3000 IU/kg of dietary vitamin D₃ (A. Schutkowski et al., 2013). The amount of vitamin D₂ in shiitake mushrooms (*Lentinula edodes*) can achieve, under optimal conditions of UV irradiation, $29.87 \pm 1.38 \mu\text{g per g dry weight}$. In the USA, Ireland, the Netherlands and Australia, fresh mushrooms are exposed to UV irradiation, which leads to an increase in the vitamin D₂ content to 10 $\mu\text{g}/100 \text{ g wet weight}$ (O. Taofiq et al., 2017; G. Cardwell et al., 2018). This is 50-100 % of the recommended daily consumption of the vitamin. The processing of baking yeast *Saccharomyces cerevisiae* by ultraviolet irradiation induces the conversion of ergosterol into vitamin D₂. The average content of vitamin D₂ is 3,065,417 IU/100 g (2,560,000-3,750,000 IU/100 g) or 770 $\mu\text{g/g}$ (640-940 $\mu\text{g/g}$), which increases its initial concentration (less than 20 IU of vitamin D₂/100 g) almost 30-50-fold (EFSA, 2014). The vitamin D₂-enriched UV-treated yeast is allowed by The European Food Safety Authority (EFSA) for fortification of yeast-leavened bread, rolls and fine pastry at maximum D₂ dose of 5 $\mu\text{g per } 100 \text{ g}$ of the products. The concentration of vitamins D₂ and D₃ after UV irradiating of the wheat germ oil (1.6 mm oil layer) was 1035 and 37 ng/g, respectively (A.C. Baur et al., 2016). Similarly, there is an increase of the vitamin D content in eggs after exposure of chickens to UV irradiation or natural sunlight (A. Schutkowski et al., 2013; J. Kühn et al., 2014, 2015). In the conditions of the complete absence of the commercial production of vitamins in our country, bio-addition with vitamin D of chicken meat, eggs and dairy products by UV irradiation of animals, mushrooms, yeast, vegetable oils takes on particular significance.

Keywords: vitamin D, biofortification, bio-addition, poultry, eggs, cows' milk, mushrooms, vitamin D-enriched UV-treated baker's yeast, ultraviolet light irradiation, wheat germ oil

Functional products for healthy lifestyle and prevention of diseases caused by an inadequate and unbalanced diet are one of the main challenges and current global trends. Comparing the people's actual diet and the state of health indicates that the diet adequate for energy expenditure does not meet the needs in macro-

and micronutrients and minor biologically active substances. The quality of a functional food product (FFP) and the effects of its systematic consumption crucially depend on the food raw materials used. Vitamin value is the most important indicator to characterize the usefulness of food raw materials and the resulting FFPs, which determines the relevance of research aimed at increasing the concentration of vitamins in farm products. To improve vitamin availability, vitamins are added directly to foods or to raw foodstuffs [1]. However, as food fortification is still a voluntary initiative of producers, the portion of such products in the retail network is low. In addition, there is unfortunately a misconception among the public and sometimes in scientific literature that synthetic vitamins are poorly assimilated by the body.

Biofortification is a modern intensively developing technology to improve composition of biologically significant nutrients in foods. In animal and poultry production, this is achieved by adding vitamins, minerals, polyunsaturated fatty acids to feed [2], and in crop production by traditional and marker-assisted selection, biotechnology and agrotechnology [3]. The promising version of biofortification is based on the ability of living organisms to form vitamin D from endogenous ergosterol or 7-dehydrocholesterol under UV radiation. Vitamin D, which is synthesized in animals, mushrooms or yeast, undergoes the stages of biotransformation and is consumed by humans in its natural form.

This overview summarizes the opportunities for increasing the vitamin D content of agricultural products through ultraviolet irradiation (UV irradiation).

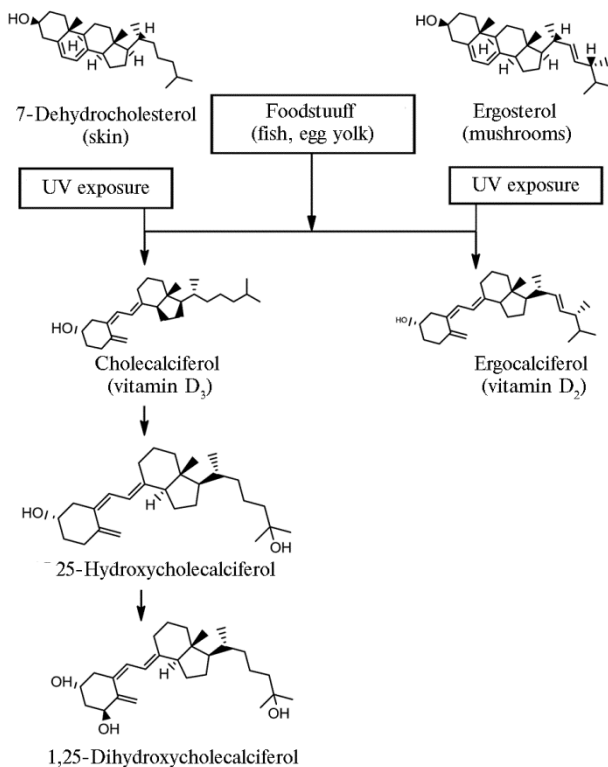
The essentiality of vitamin D (as well as other vitamins) for the human body is beyond doubt. Vitamins D, or calciferols, are represented by two compounds, the cholecalciferol (vitamin D₃) and ergocalciferol (vitamin D₂). The difference between vitamin D and other vitamins is that it not only enters the body with food but can also be formed in the skin due to ultraviolet radiation (Fig.) [4, 5].

The main sources of vitamin D₃ in the human diet (in decreasing order of content) are cod liver, fish, eggs, liver, and butter [6]. In addition to cholecalciferol, 25-hydroxy-cholecalciferol (25OHD₃) contributes significantly to the vitamin value of meat and dairy products. In food of plant origin (algae, leaves and fruits of some plants), the vitamin D amounts are extremely low (from 0.03 to 0.67 µg/100 g dry matter) [7]. Vitamin D₂ is present in mushrooms. Vitamin D vitamers have different biological activity for humans. Vitamin D₃ is more effective than D₂ [8, 9]. It was found that in tackling inadequate vitamin D status 10 µg of vitamin D₃ is equivalent to 23 µg of vitamin D₂ or 6.8 µg of 25OHD₃ [10, 11].

Calciferols are formed due to photo-isomerization of provitamins under UV radiation (see Fig.). Terrestrial animals are able to synthesize 7-dehydrocholesterol, the provitamin of cholecalciferol, from cholesterol; mushrooms and yeasts contain ergosterol, the provitamin of ergocalciferol [5]. Cholecalciferol is formed from 7-dehydrocholesterol when irradiated with sunlight or artificial ultraviolet light ($\lambda = 280-320$ nm) as a result of photochemical modification of 7-dehydrocholesterol followed by non-enzymatic isomerization.

It is believed that up to 80% of vitamin D can be synthesized in human skin under the influence of B ultraviolet radiation ($\lambda = 290-315$ nm) [12]. However, in Russia, due to insufficient sun exposure, endogenous vitamin D synthesis in the skin does not meet the body's need for this vitamin.

After entering the body, vitamin D is hydroxylated (see Fig.), turning first into 25-hydroxyvitamin D (25OHD) circulating in the blood (blood 25OHD is measured to assess individual vitamin D status) and then into its metabolically active form, the 1,25-dihydroxyvitamin D, possessing hormonal function [12].



UV dependent endogenous vitamin D synthesis and metabolism as adapted from [39].

lation, vitamin D deficiency or insufficient intake occur in 50-90% of the adults and children [4, 14, 20, 21]. This problem is solved in some countries (USA, UK, Germany, Italy, Belgium, Finland) by legally regulated technological food fortification of mass consumption products [22]. As a result, technologically enriched liquid dairy products contribute 28-63% of vitamin D [23] to the total consumption. The effectiveness of such enrichment has been proven. In particular, there are reports on a decrease in the osteoporotic fractures [24, 25] and economic benefits [26, 27]. An alternative way to increase the vitamin D content of agricultural products is to irradiate them with ultraviolet light.

Cow's milk. Vitamin D content in milk depends on the intake of ergocalciferol with mushrooms and cholecalciferol from vitamin supplements, as well as endogenous synthesis in the skin [28]. Experiments using blankets and udder covers have shown that, despite the wool cover of the skin, vitamin D synthesis in cows is carried out evenly over the entire body surface [29]. Seasonal variations in the vitamin D content in cow's milk are well known, which are minimal in winter (0-0.04 µg/g fat) and maximum in summer (up to 0.014 µg/g fat) during intensive insolation at pasture [30]. Natural insolation is more effective in improving the vitamin D status in animals than the addition of vitamin D₃ or D₂ to feed [29, 31]. Artificial UV irradiation of cows for 24 days, imitating 1, 2, 3 or 4 hours of sun exposure in summer at 56° N, resulted in an increase in vitamin D₃ and 25OHD₃ content in milk [30, 32]. Such irradiation minimizes seasonal variations in vitamin D concentration in cow's milk.

Animal meat. These products have traditionally not been considered an important source of vitamin D for humans, but in recent years there have been reports of the possibility of meat intravital modification.

Vitamin D deficiency in the population, caused by inadequate food intake and/or reduced endogenous synthesis in the skin due to insolation deficiency, has highly undesirable health effects [1, 13, 14], associated not only with skeletal but also with non-skeletal functions [15]. Vitamin D deficiency correlates with many chronic age-borne diseases [16], cardiovascular disorders [17], myocardial infarction, type 2 diabetes, tuberculosis, bronchial asthma, atopic dermatitis, urticaria, cancer of the prostate, breast, intestines, autoimmune diseases [17], and is accompanied by neurocognitive disorders, depressive states, reproductive dysfunction [12, 18, 19].

According to the results of studies of vitamin supply in the Russian population,

Wavelength $\lambda = 296$ nm was optimal for the endogenous synthesis of vitamin D₃ in the pig skin. The maximum dose of 20 kJ/m² provides 3.5-4.0 μg of vitamin D₃/cm² of skin [33]. Exposure to sunlight in pigs has been found to increase the vitamin D content in the sirloin as well [34]. Exposure to sunlight increased ($p = 0.003$) the content of 25OHD, the form more effective for human, in muscle tissue and subcutaneous fat, but did not affect the amount of vitamin D₃ ($p = 0.56$). In pigs, after 14-day stay in the sun for 1 hour in summer afternoon, the content of vitamin D₃ in meat increased ($p < 0.001$) to $0.716 \pm 0.097 \mu\text{g}/100 \text{ g}$ ($28.6 \pm 3.9 \text{ MU}/100 \text{ g}$), significantly exceeding the same indicator in the control group ($0.218 \pm 0.024 \mu\text{g}/100 \text{ g}$ or $8.7 \pm 1.0 \text{ MU}/100 \text{ g}$) [34]. The amount of 25OHD₃ reached $0.281 \pm 0.014 \mu\text{g}/100 \text{ g}$ vs. $0.130 \pm 0.016 \mu\text{g}/100 \text{ g}$. The total vitamin D content (D₃ + 25OHD₃) in the sirloin of pigs exposed to UV radiation increased 2.9-fold ($0.997 \pm 0.094 \mu\text{g}/100 \text{ g}$ vs. $0.348 \pm 0.027 \mu\text{g}/100 \text{ g}$, $p = 0.001$). The fact that the vitamin D₃ content in subcutaneous fat tissue did not change under sun exposure and was the same in pigs exposed to sunlight and in animals in the control group (with a 2-fold increase in the amount of 25OHD₃), has not been explained [34]. Sun exposure has led to better vitamin D availability in pigs, as per blood 25OHD₃ level, even with sufficient vitamin D content in the diet [35]. Daily UV irradiation of mini pigs (corresponding to sun exposure at noon for 10-20 minutes) stimulated the skin synthesis of vitamin D₃ and resulted in an increase in the amount of vitamin D₃ and 25OHD₃ in blood and the carcass. The vitamin D₃ concentration in adipose tissue of mini pigs was 150-260 ng/g if the animals were exposed to UV light, and 90-150 ng/g after D₃ oral administration at a dose of up to 60 μg per day, or 3.7-4.4 $\mu\text{g}/\text{kg}$ body-weight [36]. That is, UV irradiation was more effective. Based on the data obtained, it was concluded that changing the conditions for raising pigs when animals are allowed to be in the sun provides an effective natural increase in the vitamin D content of pork products.

Similar data were obtained for other animal species. Insolation improves the vitamin D supply in cattle. Thus, in late summer and autumn, the blood concentration of 25OHD in calf was significantly higher than in early June (55.2-63.8 vs. 26.3 ng/ml) [37]. Exposure to UV B increased the amount of vitamin D in the muscular tissue of chickens 4 times more effectively than feeding them with a diet with maximum permissible vitamin D₃ level. UV exposure increased the amount of vitamin D in chicken meat from 0.16 to 0.96 $\mu\text{g}/100 \text{ g}$, even under 3,000 MU of vitamin D₃/kg feed [38].

Mushrooms. Many mushroom species contain high concentrations of ergosterol, which is transformed into vitamin D₂ under the UV radiation. Vitamin D₂ formation is influenced by temperature, humidity, UV spectrum (B or C), duration and dose of exposure [39-41]. The table shows the vitamin D₂ content of UV-irradiated freshly picked and freeze-dried mushrooms.

Vitamin D₂ content ($\mu\text{g}/\text{g}$ of dry weight) after UV irradiation of fresh and freeze-dried mushrooms ($M \pm \text{SEM}$) [42-44]

Mushrooms	Fresh	Freeze-dried and crushed
Shiitake (<i>Lentinus edodes</i>)	29.46 \pm 2.21	60
Champignon (<i>Agaricus bisporus</i>)	3.55 \pm 0.11	119
Oyster mushroom (<i>Pleurotus ostreatus</i>)	58.96 \pm 1.15	34.6

The amount of vitamin D₂ under optimal conditions of UV irradiation of shiitake mushrooms (*Lentinula edodes*) was $29.87 \pm 1.38 \mu\text{g}/\text{g}$ dry weight [45]. Drying Shiitake mushrooms in the sun increased their vitamin D₂ content 16 times, from 100 to 1,600 MU per 100 g [15]. The original method of obtaining a functional food component was to obtain a shiitake mushroom extract with an ergoste-

rol content of 15%, and then convert it to vitamin D₂ under the influence of UV radiation at $\lambda = 254$ nm [6]. Freeze-dried shiitake extract after UV irradiation contained about 37 $\mu\text{g/g}$ of vitamin D₂, which was more than 6 times its original amount. The extract can be used as a dietary supplement to food, 0.4 g of which fully meets the daily requirement for vitamin D [46]. The vitamin D₂ content in the fruitbody of six fungal genera (*Agaricus*, *Agrocybe*, *Auricularia*, *Hypsizygus*, *Lentinula* and *Pholiota*) and five species of *Pleurotus* irradiated with ultraviolet light for 2 hours increased significantly from 0-3.93 to 15.06-208.65 $\mu\text{g/g}$ [47]. The maximum content (204.7 $\mu\text{g/g}$) was in oyster mushrooms. Drying of pre-cut mushrooms significantly increased the amount of vitamin D₂ produced during subsequent UV treatment, i.e. up to 406 vs. 45 $\mu\text{g/g}$ of whole mushrooms [48]. Irradiation of dry white mushroom powder at room temperature for about 10 minutes resulted in an increase in the vitamin D₂ content to 741.50 ± 23.75 $\mu\text{g/g}$ [41].

Fresh mushrooms are currently exposed to UV radiation in the United States, Ireland, the Netherlands, and Australia, which leads to an increase in the vitamin D₂ content of up to 10 $\mu\text{g}/100$ g of raw weight [43, 49]. As a result, a portion of mushrooms (100 g) provides 50-100% of the recommended vitamin norm. The vitamin D content of the mushrooms treated in this way is similar to that of fatty fish [50]. This is essential for vegetarians typically lacking in vitamin D due to refusal of animal food [51-53]. The vitamin D₂ content in mushrooms with different methods of preparation is 62-88% of the original ($p \leq 0.05$) [54].

In rats with experimental vitamin D deficiency it was shown that vitamin D₂ from UV irradiated fungi is digested by these animals: the blood amounts of 25OHD and calcium increased and the mineral density of bone tissue was significantly higher ($p < 0.01$) than in the control rats receiving conventional mushrooms [55].

As per estimates of the vitamin D bioavailability from mushrooms, its absorption is sufficiently high. It was shown that consumption of mushrooms after their UV irradiation led to an increase in the blood concentration of 25OHD₂ in human up to 24.2 nmol/l, especially in case of initial lack of this vitamin. However, there was a 12.6 nmol/l decrease in 25OHD₃ [44]. Vitamin D₂ is well absorbed from UV irradiated shredded chanterelles and porcini mushrooms, helps to increase the 25OHD₂ concentration and reduce the amount of 25OHD₃ in blood [56]. It has been found that 2,000 MU of vitamin D₂ contained in mushrooms is as effective in increasing and maintaining the required amount of 25OHD in human blood as 2,000 MU of vitamin D₂ or D₃ [57].

A 30-day feeding calves with vitamin D₂-enriched mushrooms prior to slaughter resulted in an increase in the vitamin D content of the meat, although less pronounced than that of the diet with vitamin D₃ [58].

Bakery yeast. The UV irradiation on *Saccharomyces cerevisiae* bakery yeast has induced the conversion of the ergosterol they contain into vitamin D₂ (see Fig.). The amount of vitamin D₂ was almost 30-50 times higher than its original concentration (less than 20 MU/100 g) and averaged 3,065 thousand MU/100 g (fluctuations within the range of 2,560-3,750 thousand MU/100 g), or 770 $\mu\text{g/g}$ (640-940 $\mu\text{g/g}$) [59]. In 2012, the European Food Safety Authority (EFSA, Italy) approved UV-treated yeast enriched with vitamin D₂ as a new food ingredient in the production of yeast bread, rolls, flour confectionery products, the maximum dose is 5 $\mu\text{g}/100$ g. Such yeast is also permitted as a component of dietary supplements.

Consumption of bread made with D₂-rich yeast had a comparable effect with pure vitamin D₂ on the 25OHD concentration in women blood plasma [59]. The vitamin D₂ from UV-irradiated yeast used in bread baking was shown to be digestible and improve bone health in rats with an initial vitamin D defi-

ciency [60]. According to the effect on the content of 25OHD in experiments on growing rats, the effectiveness of the used form of vitamin D decreased in the series vitamin D₃ > vitamin D₂ > UV-treated yeast cells or their individual fractions [61].

In Canada, vitamin D₂ containing yeast are allowed as an ingredient of bread products in quantities up to 90 MU (2.25 µg) per 100 g, which is 23% of the D₂ recommended consumption rate [62]. In the United States, legislation was amended in 2012 to ensure the safe use of vitamin D₂ from baking yeast in baking products: it is permissible to add no more than 400 MU of vitamin D₂ per 100 g of finished products, or 50% of the recommended intake [63]. Food yeast remains a potential source of biologically active substances [64].

Vegetable oils. Vegetable oils can also be a potential source of vitamin D. They contain significant amounts of not only ergosterol but also 7-dehydrocholesterol (7-DHC) [65]. The ergosterol concentration is 22.1-34.2 µg/g in wheat germ oil, 4.2-23.4 µg/g in avocado oil, 7.9-17.4 µg/g in sunflower oil, 4.1-9.5 µg/g in rapeseed, soybean, flax oil, and < 4.5 µg/g in olive oil. UV-exposure resulted in the partial conversion of ergosterol and 7-DHC to vitamins D₂ and D₃ in these oils [65]. After 1-minute UV irradiation of 1.6 mm layer of wheat germ oil, the concentration of vitamins D₂ and D₃ was 1035 and 37 ng/g, respectively [65]. At the same time, such influence practically did not reduce the content of tocopherols and did not intensify peroxidation [65]. Assessment of the bioavailability of vitamin D showed that mice with an initial deficit of vitamin D improved, which was confirmed by an increase in the blood 25OHD concentration and accumulation in the liver compared to mice that received conventional wheat germ oil. However, the plasma content of 25OHD in mice fed UV-treated oil did not reach the values observed in the group that consumed oil with pure vitamin D added [65].

Chicken egg. Enrichment of eggs with vitamin D instead of adding it to the feed of chickens can be achieved by artificially irradiating birds with ultraviolet (bio-addition), or through free-range poultry.

In eggs from laying hens irradiated with ultraviolet for 3 hours daily for 4 weeks vs. those fed diet with the adequate vitamin D₃ content (3,000 MU/kg feed), the amount of vitamin D (cholecalciferol and 25OHD) usually reaches 2.5 µg. It is almost 5 times higher than in eggs of hens on the same diet but not UV irradiated [38]. Curiously, the endogenous formation of the vitamin occurs mainly in the legs of chickens, where the plumage is the smallest. The dependence of the increase in the vitamin D content in the egg yolk on the time of daily UV radiation is non-linear. With daily exposure to UV light for 300 minutes, the vitamin D₃ content increased to 28.6 µg/100 g of egg yolk dry weight, but did not reach the plateau, while the amount of 25OHD was already maximum with exposure for 60 min [66].

A similar increase in the vitamin D content is due to natural insolation. The amount of vitamin D₃ in the egg yolk in birds exposed to sunlight (free and closed/free keeping) was 3-4 times higher ($p < 0.001$) than its accumulation in the egg yolk of chickens in closed range [67]. The concentration of vitamin D₃ in the egg yolk of birds in free range was 14.3 µg/100 g vs. 3.8 µg/100 g of dry weight. The vitamin D content in the egg yolk of chicken eggs under the mixed keeping mode is in an intermediate position. The amount of 25OHD₃ in the egg yolk also depended on sunlight, although it was lower than the concentration of vitamin D₃ ($p < 0.05$).

It should be noted that in recent years there has been a kind of rehabilitation of the chicken egg. Consumption of 6 to 12 eggs per week in balanced diets has no negative effect on major risk factors for cardiovascular disease and

type 2 diabetes [68]. A nutritional analysis of 7,216 participants aged 55-80 found that moderate egg intake was not associated with increased risk of cardiovascular disease in both diabetic and non-diabetic patients [69]. In addition, the simultaneous consumption of whole eggs cooked with fresh vegetable salad is an effective way to increase the absorption of α -tocopherol and γ -tocopherol, as well as carotenoids from plant foods [70, 71].

An adequate supply of vitamin D₃ is essential to maintain public health. In recent years, many countries have applied technological fortification of food products (yoghurt, bread, etc.) [27, 72, 73]. As to the role of biofortification, it should be noted that there is no production of vitamin substances in Russia. Food, medical and agricultural requirements for vitamin substances are met only through imports [72].

Thus, the use of alternative ways of enriching food products with vitamin D is vital, to some extent contributing to the solution of the problem of import substitution. Biofortification of chicken, pork, eggs and dairy products with vitamin D by UV irradiation of animals is perspective. So far, plant sources of vitamin D have not been given due importance, but the possibility of increasing the vitamin D content of mushrooms and plant oils through UV irradiation, makes it advisable to obtain these products that are important for vegetarians.

REFERENCES

1. Kodentsova V.M., Risnik D.V. *Voprosy dietologii*, 2017, 7(2): 33-40 (doi: 10.20953/2224-5448-2017-2-33-40) (in Russ.).
2. Kavtarashvili A.Sh., Mazo V.K., Kodentsova V.M., Risnik D.V., Stefanova I.L. Biofortification of hen eggs: vitamins and carotenoids (review). *Sel'skokhozyaistvennaya biologiya [Agricultural Biology]*, 2017, 52(6): 1094-1104 (doi: 10.15389/agrobiology.2017.6.1094eng).
3. Garg M., Sharma N., Sharma S., Kapoor P., Kumar A., Chunduri V., Arora P. Biofortified crops generated by breeding, agronomy, and transgenic approaches are improving lives of millions of people around the world. *Front. Nutr.*, 2018, 5: 12 (doi: 10.3389/fnut.2018.00012).
4. Kodentsova V.M., Mendel' O.I., Khotimchenko S.A., Baturin A.K., Nikityuk D.B., Tutel'yan V.A. *Voprosy pitaniya*, 2017, 86(2): 47-62 (doi: 10.24411/0042-8833-2017-00067) (in Russ.).
5. Göring H. Vitamin D in nature: a product of synthesis and/or degradation of cell membrane components. *Biochemistry (Moscow)*, 2018, 83(11): 1350-1357 (doi: 10.1134/S0006297918110056).
6. Schmid A., Walther B. Natural vitamin D content in animal products. *Advances in Nutrition*, 2013, 4(4): 453-462 (doi: 10.3945/an.113.003780).
7. Hughes L.J., Black L.J., Sherriff J.L., Dunlop E., Strobel N., Lucas R.M., Bornman J.F. Vitamin D content of Australian native food plants and Australian-grown edible seaweed. *Nutrients*, 2018, 10(7): 876 (doi: 10.3390/nu10070876).
8. Guo J., Lovegrove J.A., Givens D.I. 25(OH)D₃-enriched or fortified foods are more efficient at tackling inadequate vitamin D status than vitamin D₃. *Proceedings of the Nutrition Society*, 2018, 77(3): 282-291 (doi: 10.1017/S0029665117004062).
9. Tripkovic L., Lambert H., Hart K., Smith C.P., Bucca G., Penson S., Chope G., Hypponen E., Berry J., Vieth R., Lanham-New S. Comparison of vitamin D₂ and vitamin D₃ supplementation in raising serum 25 hydroxyvitamin D status: a systematic review and meta analysis. *The American Journal of Clinical Nutrition*, 2012, 95(6): 1357-1364 (doi: 10.3945/ajcn.111.031070).
10. Jakobsen J., Andersen E., Christensen T., Andersen R., Bügel S. Vitamin D vitamers affect vitamin D status differently in young healthy males. *Nutrients*, 2018, 10(1): 2 (doi: 10.3390/nu10010012).
11. Wilson L.R., Tripkovic L., Hart K.H., Lanham-New S.A. Vitamin D deficiency as a public health issue: using vitamin D₂ or vitamin D₃ in future fortification strategies. *Proceedings of the Nutrition Society*, 2017, 76(3): 392-399 (doi: 10.1017/S0029665117000349).
12. Hossein-nezhad A., Holick M.F. Vitamin D for health: A global perspective. *Mayo Clinic Proceedings*, 2013, 88(7): 720-755 (doi: 10.1016/j.mayocp.2013.05.011).
13. Kodentsova V.M., Risnik D.V. V sbornike: *Ekologiya. Ekonomika. Informatika. Tom 1: Sistemnyi analiz i modelirovanie ekonomicheskikh i ekologicheskikh system* [In: Ecology. Economy. Computer science. Vol. 1: System analysis and modeling of economic and environmental systems]. Rostov-na-Donu, 2016: 486-498 (in Russ.).
14. Zakharova I.N., Mal'tsev S.V., Borovik T.E., Yatsyk G.V., Malyavskaya S.I., Vakhlova I.V., Shumatova T.A., Romantsova E.B., Romanyuk F.P., Klimov L.Ya., Pirozhkova N.I., Kolesni-

- kova S.M., Ku'ryaninova V.A., Tvorogova T.M., Vasil'eva S.V., Mozhukhina M.V., Evseeva E.A. *Pediatrics. Zhurnal im. G.N. Speranskogo*, 2015, 94(1): 62-67 (in Russ.).
15. Wacker M., Holick M.F. Vitamin D — effects on skeletal and extraskeletal health and the need for supplementation. *Nutrients*, 2013, 5(1): 111-148 (doi: 10.3390/nu5010111).
 16. Drapkina O.M., Shepel' R.N., Fomin V.V., Svistunov A.A. *Terapevticheskii arkhiv*, 2018, 90(1): 69-75 (in Russ.).
 17. Podzolkov V.I., Pokrovskaya A.E., Panasenko O.I. *Terapevticheskii arkhiv*, 2018, 90(9): 144-150 (in Russ.).
 18. Kalinchenko S.Yu., Korotkova N.A. *Voprosy dietologii*, 2018, 8(2): 32-37 (doi: 10.20953/2224-5448-2018-2-32-37) (in Russ.).
 19. Zaidieva Ya.Z. *Sovremennaya ginekologiya*, 2018, 1(6): 24-33 (in Russ.).
 20. Kodentsova V.M., Beketova N.A., Nikityuk D.B., Tutel'yan V.A. *Profilakticheskaya meditsina*, 2018, 21(4): 32-37 (doi: 10.17116/profmed201821432) (in Russ.).
 21. Zakharova I.N., Tvorogova T.M., Gromova O.A., Evseeva E.A., Lazareva S.I., Maikova I.D., Sugyan N.G. *Pediatriceskaya farmakologiya*, 2015, 12(5): 528-531 (doi: 10.15690/pf.v12i5.1453) (in Russ.).
 22. Kodentsova V.M., Vrzhesinskaya O.A. *Voprosy pitaniya*, 2016, 85(2): 31-50 (in Russ.).
 23. Itonen S.T., Erkkola M., Lamberg-Allardt C.J.E. Vitamin D fortification of fluid milk products and their contribution to vitamin D intake and vitamin D status in observational studies — a review. *Nutrients*, 2018, 10(8): 1054 (doi: 10.3390/nu10081054).
 24. Hilgsmann M., Bulet N., Fardellone P., Al-Daghri N., Reginster J.-Y. Public health impact and economic evaluation of vitamin D-fortified dairy products for fracture prevention in France. *Osteoporosis International*, 2017, 28(3): 833-840 (doi: 10.1007/s00198-016-3786-1).
 25. Raulio S., Erlund I., Männistö S., Sarlio-Lähteenkorva S., Sundvall J., Tapanainen H., Virtanen S.M. Successful nutrition policy: improvement of vitamin D intake and status in Finnish adults over the last decade. *European Journal of Public Health*, 2016, 27(2): 268-273 (doi: 10.1093/eurpub/ckw154).
 26. Jääskeläinen T., Itonen S.T., Lundqvist A., Erkkola M., Koskela T., Lakkala K., Dowling K.G., Hull G.L., Kröger H., Karppinen J., Kyllönen E., Härkänen T., Cashman K.D., Männistö S., Lamberg-Allardt C. The positive impact of general vitamin D food fortification policy on vitamin D status in a representative adult Finnish population: evidence from an 11-y follow-up based on standardized 25-hydroxyvitamin D data. *The American Journal of Clinical Nutrition*, 2017, 105(6): 1512-1520 (doi: 10.3945/ajcn.116.151415).
 27. Pilz S., März W., Cashman K.D., Kiely M.E., Whiting S.J., Holick M.F., Grant W.B., Pludowski P., Hilgsmann M., Trummer C., Schwetz V., Lerchbaum E., Pandis M., Tomaschitz A., Gröbler M.R., Gaksch M., Verheyen N., Hollis B.W., Rejnmark L., Karras S.N., Hahn A., Bischoff-Ferrari H.A., Reichrath J., Jorde R., Elmadfa I., Vieth R., Scragg R., Calvo M.S., van Schoor N.M., Bouillon R., Lips P., Itonen S.T., Martineau A.R., Lamberg-Allardt C., Zittermann A. Rationale and plan for vitamin D food fortification: a review and guidance paper. *Front. Endocrinol.*, 2018, 9: 373 (doi: 10.3389/fendo.2018.00373).
 28. Hymøller L., Jensen S.K. Plasma transport of ergocalciferol and cholecalciferol and their 25-hydroxylated metabolites in dairy cows. *Domestic Animal Endocrinology*, 2017, 59: 44-52 (doi: 10.1016/j.domaniend.2016.11.002).
 29. Hymøller L., Jensen S.K. Vitamin D₃ synthesis in the entire skin surface of dairy cows despite hair coverage. *Journal of Dairy Science*, 2010, 93(5): 2025-2029 (doi: 10.3168/jds.2009-2991).
 30. Weir R.R., Strain J.J., Johnston M., Lewis C., Fearon A.M., Stewart S., Pourshahidi L.K. Environmental and genetic factors influence the vitamin D content of cows' milk. *Proceedings of the Nutrition Society*, 2017, 76(1): 76-82 (doi: 10.1017/S0029665116000811).
 31. Yue Y., Hymøller L., Jensen S.K., Lauridsen C. Effect of vitamin D treatments on plasma metabolism and immune parameters of healthy dairy cows. *Archives of Animal Nutrition*, 2018, 72(3): 205-220 (doi: 10.1080/1745039X.2018.1448564).
 32. Jakobsen J., Jensen S.K., Hymøller L., Andersen E.W., Kaas P., Burild A., Jäpelt R.B. Short communication: artificial ultraviolet B light exposure increases vitamin D levels in cow plasma and milk. *Journal of Dairy Science*, 2015, 98(9): 6492-6498 (doi: 10.3168/jds.2014-9277).
 33. Barnkob L.L., Argyraki A., Petersen P.M., Jakobsen J. Investigation of the effect of UV-LED exposure conditions on the production of vitamin D in pig skin. *Food Chemistry*, 2016, 212: 386-391 (doi: 10.1016/j.foodchem.2016.05.155).
 34. Larson-Meyer D.E., Ingold B.C., Fensterseifer S.R., Austin K.J., Wechsler P.J., Hollis B.W., Makowski A.J., Alexander B.M. Sun exposure in pigs increases the vitamin D nutritional quality of pork. *PLoS ONE*, 2017, 12(11): e0187877 (doi: 10.1371/journal.pone.0187877).
 35. Alexander B.M., Ingold B.C., Young J.L., Fensterseifer S.R., Wechsler P.J., Austin K.J., Larson-Meyer D.E. Sunlight exposure increases vitamin D sufficiency in growing pigs fed a diet formulated to exceed requirements. *Domestic Animal Endocrinology*, 2017, 59: 37-43 (doi: 10.1016/j.domaniend.2016.10.006).

36. Burild A., Frandsen H.L., Poulsen M., Jakobsen J. Tissue content of vitamin D₃ and 25-hydroxy vitamin D₃ in minipigs after cutaneous synthesis, supplementation and deprivation of vitamin D₃. *Steroids*, 2015, 98: 72-79 (doi: 10.1016/j.steroids.2015.02.017).
37. Casas E., Lippolis J.D., Kuehn L.A., Reinhardt T.A. Seasonal variation in vitamin D status of beef cattle reared in the central United States. *Domestic Animal Endocrinology*, 2015, 52: 71-74 (doi: 10.1016/j.domaniend.2015.03.003).
38. Schutkowski A., Krämer J., Kluge H., Hirche F., Krombholz A., Theumer T., Stangl G.I. UVB exposure of farm animals: study on a food-based strategy to bridge the gap between current vitamin D intakes and dietary targets. *PLoS ONE*, 2013, 8(7): e69418 (doi: 10.1371/journal.pone.0069418).
39. Taofiq O., Fernandes B., Barros L., Barreiro M.F., Ferreira I.C. UV-irradiated mushrooms as a source of vitamin D₂: a review. *Trends in Food Science & Technology*, 2017, 70: 82-94 (doi: 10.1016/j.tifs.2017.10.008).
40. Edward T.L., Kirui M.S.K., Omolo J.O., Ngumbu R.G., Odhiambo P.M. Change in concentration of vitamin D₂ in oyster mushrooms exposed to 254nm and 365nm UV-light during growth. *International Journal of Biochemistry and Biophysics*, 2015, 3(1): 1-5.
41. Lee N.K., Aan B.Y. Optimization of ergosterol to vitamin D₂ synthesis in *Agaricus bisporus* powder using ultraviolet-B radiation. *Food Science and Biotechnology*, 2016, 25(6): 1627-1631 (doi: 10.1007/s10068-016-0250-0).
42. Slawinska A., Fornal E., Radzki W., Jablonska-Rys E., Parfieniuk E. Vitamin D₂ stability during the refrigerated storage of ultraviolet B-treated cultivated culinary-medicinal mushrooms. *International Journal of Medicinal Mushrooms*, 2017, 19(3): 249-255 (doi: 10.1615/IntJMedMushrooms.v19.i3.70).
43. Cardwell G., Bornman J.F., James A.P., Black L.J. A review of mushrooms as a potential source of dietary vitamin D. *Nutrients*, 2018, 10(10): E1498 (doi: 10.3390/nu10101498).
44. Cashman K.D., Kiely M., Seamans K.M., Urbain P. Effect of ultraviolet light-exposed mushrooms on vitamin D status: liquid chromatography-tandem mass spectrometry reanalysis of biobanked sera from a randomized controlled trial and a systematic review plus meta-analysis. *The Journal of Nutrition*, 2016, 146(3): 565-75 (doi: 10.3945/jn.115.223784).
45. Won D.J., Kim S.Y., Jang C.H., Lee J.S., Ko J.A., Park H.J. Optimization of UV irradiation conditions for the vitamin D₂-fortified shiitake mushroom (*Lentinula edodes*) using response surface methodology. *Food Science and Biotechnology*, 2017, 27(2): 417-424 (doi: 10.1007/s10068-017-0266-0).
46. Chien R.C., Yang S.C., Lin L.M., Mau J.L. Anti-inflammatory and antioxidant properties of pulsed light irradiated *Lentinula edodes*. *Journal of Food Processing and Preservation*, 2017, 41(4): e13045 (doi: 10.1111/jfpp.13045).
47. Morales D., Gil-Ramirez A., Smiderle F.R., Piris A.J., Ruiz-Rodriguez A., Soler-Rivas C. Vitamin D-enriched extracts obtained from shiitake mushrooms (*Lentinula edodes*) by supercritical fluid extraction and UV-irradiation. *Innovative Food Science & Emerging Technologies*, 2017, 41: 330-336 (doi: 10.1016/j.ifset.2017.04.008).
48. Nolle N., Argyropoulos D., Ambacher S., Müller J., Biesalski H.K. Vitamin D₂ enrichment in mushrooms by natural or artificial UV-light during drying. *LWT - Food Science and Technology*, 2017, 85(part B): 400-404 (doi: 10.1016/j.lwt.2016.11.072).
49. Calvo M.S., Whiting S.J. Survey of current vitamin D food fortification practices in the United States and Canada. *The Journal of Steroid Biochemistry and Molecular Biology*, 2013, 136: 211-213 (doi: 10.1016/j.jsbmb.2012.09.034).
50. *Khimicheskii sostav rossiiskikh pishchevykh produktov /Pod redaktsiei I.M. Skurikhina, V.A. Tutel'yan* [Chemical composition of foodstuffs produced in Russia. I.M. Skurikhin, V.A. Tutel'yan (eds.)]. Moscow, 2002 (in Russ.).
51. Laskowska-Klita T., Chelchowska M., Ambroszkiewicz J., Gajewska J., Klemarczyk W. The effect of vegetarian diet on selected essential nutrients in children. *Medycyna Wieku Rozwojowego*, 2011, 15(3): 318-325.
52. Elorinne A.L., Alfthan G., Erlund I., Kivimäki H., Paju A., Salminen I., Turpeinen U., Voutilainen S., Laakso J. Food and nutrient intake and nutritional status of Finnish vegans and non-vegetarians. *PLoS ONE*, 2016, 11(2): e0148235 (doi: 10.1371/journal.pone.0148235).
53. Gorbachev D.O., Sazonova O.V., Gil'miyarova F.N., Gussyakova O.A., Myakisheva Yu.V., Beketova N.A., Kodentsova V.M., Vrzhesinskaya O.A., Gorbacheva I.V., Gavryushin M.Yu. *Profilakticheskaya meditsina*, 2018, 21(3): 51-56 (doi: 10.17116/profmed201821351) (in Russ.).
54. Ložnjak P., Jakobsen J. Stability of vitamin D₃ and vitamin D₂ in oil, fish and mushrooms after household cooking. *Food Chemistry*, 2018, 254: 144-149 (doi: 10.1016/j.foodchem.2018.01.182).
55. Jasinghe V.J., Perera C.O., Barlow P.J. Bioavailability of vitamin D₂ from irradiated mushrooms: an in vivo study. *British Journal of Nutrition*, 2005, 93(6): 951-955 (doi: 10.1079/BJN20051416).
56. Stephensen C.B., Zerofsky M., Burnett D.J., Lin Y.P., Hammock B.D., Hall L.M., McHugh T. Ergocalciferol from mushrooms or supplements consumed with a standard meal increases 25-hydroxyergocalciferol but decreases 25-hydroxycholecalciferol in the serum of healthy adults. *The Journal of Nutrition*, 2012, 142(7): 1246-1252 (doi: 10.3945/jn.112.159764).

57. Keegan R.-J.H., Lu Z., Bogusz J.M., Williams J.E., Holick M.F. Photobiology of vitamin D in mushrooms and its bioavailability in humans. *Dermato-Endocrinology*, 2013, 5(1): 165-176 (doi: 10.4161/derm.23321).
58. Duffy S.K., O'Doherty J.V., Rajauria G., Clarke L.C., Hayes A., Dowling K.G., O'Grady M.N., Kerry J.P., Jakobsen J., Cashman K.D., Kelly A.K. Vitamin D-biofortified beef: A comparison of cholecalciferol with synthetic versus UVB-mushroom-derived ergosterol as feed source. *Food Chemistry*, 2018, 256: 18-24 (doi: 10.1016/j.foodchem.2018.02.099).
59. EFSA NDA Panel (EFSA Panel on Dietetic Products, Nutrition and Allergies). Scientific opinion on the safety of vitamin D-enriched UV-treated baker's yeast. *EFSA Journal*, 2014, 12(1): 3520 (doi: 10.2903/j.efsa.2014.3520).
60. Hohman E.E., Martin B.R., Lachcik P.J., Gordon D.T., Fleet J.C., Weaver C.M. Bioavailability and efficacy of vitamin D₂ from UV-irradiated yeast in growing, vitamin D-deficient rats. *J. Agric. Food Chem.*, 2011, 59(6): 2341-2346 (doi: 10.1021/jf104679c).
61. Itkonen S.T., Pajula E.T., Dowling K.G., Hull G.L., Cashman K.D., Lamberg-Allardt C.J. Poor bioavailability of vitamin D₂ from ultraviolet-irradiated D₂-rich yeast in rats. *Nutrition Research*, 2018, 59: 36-43 (doi: 10.1016/j.nutres.2018.07.008).
62. Health Canada. Department of health, food and drugs regulation — Amendments. *Canada Gazette Part I*, 19 February, 2011: 439-440.
63. FDA (Food and Drug Administration). *Food and drug administration, Department of health and human services. Food additives permitted for direct addition to food for human consumption; vitamin D₂ baker's yeast. Federal Register 08/29/2012*. Available <http://federalregister.gov/a/2012-21353>. Accessed 12.08.2019.
64. Shurson G.C. Yeast and yeast derivatives in feed additives and ingredients: Sources, characteristics, animal responses, and quantification methods. *Animal Feed Science and Technology*, 2018, 235: 60-76 (doi: 10.1016/j.anifeedsci.2017.11.010).
65. Baur A.C., Brandsch C., König B., Hirche F., Stangl G.I. Plant oils as potential sources of vitamin D. *Frontiers in Nutrition*, 2016, 12(3): 29 (doi: 10.3389/fnut.2016.00029).
66. Kühn J., Schutkowski A., Hirche F., Baur A.C., Mielenz N., Stangl G.I. Non-linear increase of vitamin D content in eggs from chicks treated with increasing exposure times of ultraviolet light. *The Journal of Steroid Biochemistry and Molecular Biology*, 2015, 148: 7-13 (doi: 10.1016/j.jsbmb.2014.10.015).
67. Kühn J., Schutkowski A., Kluge H., Hirche F., Stangl G.I. Free-range farming: a natural alternative to produce vitamin D-enriched eggs. *Nutrition*, 2014, 30(4): 481-484 (doi: 10.1016/j.nut.2013.10.002).
68. Richard C., Cristall L., Fleming E., Lewis E.D., Ricupero M., Jacobs R.L., Field C.J. Impact of egg consumption on cardiovascular risk factors in individuals with type 2 diabetes and at risk for developing diabetes: a systematic review of randomized nutritional intervention studies. *Canadian Journal of Diabetes*, 2017, 41(4): 453-463 (doi: 10.1016/j.cjcd.2016.12.002).
69. Diez-Espino J., Basterra-Gortari F.J., Salas-Salvadó J., Buil-Cosiales P., Corella D., Schröder H., Estruch R., Ros E., Gómez-Gracia E., Arós F., Fiol M., Lapetra J., Serra-Majem L., Pintó X., Babio N., Quiles L., Fito M., Martí A., Toledo E. Egg consumption and cardiovascular disease according to diabetic status: The PREDIMED study. *Clinical Nutrition*, 2017, 36(4): 1015-1021 (doi: 10.1016/j.clnu.2016.06.009).
70. Kim J.E., Ferruzzi M.G., Campbell W.W. Egg consumption increases vitamin E absorption from co-consumed raw mixed vegetables in healthy young men. *The Journal of Nutrition*, 2016, 146(11): 2199-2205 (doi: 10.3945/jn.116.236307).
71. Kim J.E., Gordon S.L., Ferruzzi M.G., Campbell W.W. Effects of egg consumption on carotenoid absorption from co-consumed, raw vegetables. *The American Journal of Clinical Nutrition*, 2015, 102(1): 75-83 (doi: 10.3945/ajcn.115.111062).
72. Kodentsova V.M., Vrzhesinskaya O.A., Risnik D.V., Nikityuk D.B., Tutel'yan V.A. *Voprosy pitaniya*, 2017, 86(4): 113-124 (in Russ.).
73. Tripkovic L., Wilson L.R., Hart K., Johnsen S., de Lusignan S., Smith C.P., Hypponen E. Daily supplementation with 15 µg vitamin D₂ compared with vitamin D₃ to increase wintertime 25-hydroxyvitamin D status in healthy South Asian and white European women: a 12-wk randomized, placebo-controlled food-fortification trial. *The American Journal of Clinical Nutrition*, 2017, 106(2): 481-490 (doi: 10.3945/ajcn.116.138693).